

Forum Review

Activation of Tyrosine Kinases by Reactive Oxygen Species in Vascular Smooth Muscle Cells: Significance and Involvement of EGF Receptor Transactivation by Angiotensin II

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ABSTRACT

Enhanced production of reactive oxygen species (ROS) such as H_2O_2 and a failure in ROS removal by scavenging systems are hallmarks of several cardiovascular diseases such as atherosclerosis and hypertension. ROS act as second messengers that play a prominent role in intracellular signaling and cellular function. In vascular smooth muscle cells (VSMCs), a vascular pathogen, angiotensin II, appears to initiate growth-promoting signal transduction through ROS-sensitive tyrosine kinases. However, the precise mechanisms by which tyrosine kinases are activated by ROS remain unclear. In this review, the current knowledge that suggests how certain tyrosine kinases are activated by ROS, along with their functional significance in VSMCs, will be discussed. Recent findings suggest that transactivation of the epidermal growth factor receptor by ROS requires metalloprotease-dependent heparin-binding epidermal growth factor-like growth factor production, whereas other ROS-sensitive tyrosine kinases such as PYK2, JAK2, and platelet-derived growth factor receptor require activation of protein kinase C- δ . Each of these ROS-sensitive kinases could mediate specific signaling critical for pathophysiological responses. Detailed analysis of the mechanism of cross-talk and the downstream function of these various tyrosine kinases will yield new therapeutic interventions for cardiovascular disease. *Antioxid. Redox Signal.* 5, 771–780.

INTRODUCTION

INCREASING EVIDENCE over the past several years has revealed important clues regarding reactive oxygen species (ROS), in particular, their role in both normal and abnormal cellular function. ROS are reduction/oxidation molecules derived from molecular oxygen that include superoxide anion ($O_2^{\bullet-}$), hydrogen peroxide (H_2O_2), and hydroxyl radical (OH^\bullet). They are generated by a variety of extracellular stimuli such as growth factors, G protein-coupled receptor (GPCR) agonists, cytokines, ultraviolet radiation, increased osmolarity, and other cellular stresses (22, 45). Recently, ROS were widely recognized as prominent players in the pathophysiology of many diseases, including cardiovascular diseases such as hypertension, atherosclerosis, and restenosis after vascular injury (8,

32). In fact, the majority of vascular cell types that construct vessel walls generate ROS, supporting the notion that ROS may function as regulators and inducers of these diseases (32). Currently, ROS are proposed to induce cardiovascular diseases by three major mechanisms: (1) oxidation of lipid (LDL) to produce oxidized LDL, which plays a major role in the development of atherosclerosis; (2) inhibition of nitric oxide function through $O_2^{\bullet-}$ and nitric oxide interaction that forms peroxynitrite; and (3) activation of intracellular signal transduction pathways as second messengers (8, 32, 50).

ROS-mediated signaling pathways are known to promote a variety of cellular events, such as growth, differentiation, survival, apoptosis, and inflammation, as well as gene expression, in vascular cells (32, 38, 50). However, of particular interest is the precise mechanism of how ROS stimulate or modulate

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extracellular or intracellular targets that leads to pathophysiological responses in vascular cells. Phosphorylation of receptor and non-receptor tyrosine kinases is a critical step in signal transduction that leads to specific cellular functions such as cell growth. Interestingly, both protein tyrosine phosphorylation and ROS production are commonly induced by several growth factors, and ROS can activate many tyrosine kinases. Thus, understanding the mechanism and downstream consequences of ROS-mediated tyrosine kinase activation is a current research focus by many researchers. Researchers have elucidated important roles and mechanisms of ROS in vascular smooth muscle cells (VSMCs), especially in regards to activation of tyrosine kinases, which could be responsible for specific cellular responses leading to cardiovascular diseases.

It is now recognized that events such as hypertrophy, hyperplasia, and migration of VSMCs may be initiated through tyrosine kinase activation induced by ROS production in response to various vascular pathogens such as angiotensin II (AngII) and platelet-derived growth factor (PDGF). AngII, a highly studied vasoactive hormone of the renin-angiotensin system, plays a significant role in the regulation of cardiovascular homeostasis mainly through the AngII type 1 (AT₁) receptor (90). One major undesired consequence of AngII is its role in the development of cardiovascular diseases, including hypertension, atherosclerosis, and heart failure (13, 14, 29, 79). Stimulated AT₁ receptors lead to rapid protein tyrosine phosphorylation of various signaling molecules in AngII target cells, and thereby lead to a prototypical growth promoting signal such as mitogen-activated protein kinase (MAPK/extracellular signal-regulated kinase [ERK]) activation and induction of *c-fos* and *c-jun* expression (7, 15, 31). ROS have been shown to mediate many of the pathological effects of AngII. In VSMCs, AngII is a potent ROS inducer that generates the majority of ROS through NAD(P)H oxidase. Acting via a ROS-dependent mechanism, AngII mediates hypertrophy and/or proliferation of VSMCs (30).

PDGF is produced in a variety of cardiovascular cells and is one of the strongest inducers of VSMC growth and migration (35). Importantly, PDGF is also a potent inducer of ROS in VSMCs and is known to stimulate p42/44 MAPK/ERK and subsequent mitogenic responses through ROS production (86). Pathophysiologically, PDGF has been reported to play a role in cardiovascular diseases such as atherosclerosis and restenosis, which could involve ROS-dependent mechanisms (35, 36).

New accumulating evidence suggests that activation of several ROS-sensitive tyrosine kinases is an important initial sig-

naling trigger that leads to specific signaling events responsible for functional modulation of VSMCs. Therefore, in this review, the discussion will focus on current findings that suggest novel mechanisms of how ROS mediate the activation of several receptor and non-receptor tyrosine kinases in VSMCs, as well as the pathophysiological significance of their activation.

ROS-ACTIVATED TYROSINE KINASES
IN VSMCS

Tyrosine kinases (receptor and non-receptor) in VSMCs that are ROS-sensitive or utilize ROS-dependent mechanisms for activation are listed in Table 1 (21, 24, 25, 28, 33, 55, 56, 70, 71, 74, 76, 78, 84, 86, 93). It can be hypothesized that multiple risk factors → ROS → tyrosine kinase activation → abnormal VSMC response → cardiovascular disease. Generally, three mechanisms have been proposed by which ROS activate tyrosine kinases. First, ROS may directly activate kinases by altering protein-protein interactions depending on sulfhydryl groups. Second, protein tyrosine phosphatases that contain a cysteine residue in their activation site may be directly inhibited by ROS, which in turn results in tyrosine phosphorylation of the kinases and may affect their activities. Third, oxidation stimulates proteolysis of regulatory proteins that may inhibit tyrosine kinase activity (8). It has been suggested that at least two or more distinct mechanisms are involved in tyrosine kinase activation by ROS in VSMCs. In the following sections of this review, we will discuss the role of each tyrosine kinase with respect to how ROS induce their activation and the functional and pathophysiological consequence in VSMCs. In addition to the tyrosine kinases listed in Table 1, several other tyrosine kinases expressed in different cell types have also been shown to be activated via a ROS-dependent mechanism. However, the possible roles of these potential ROS-sensitive tyrosine kinases such as insulin-like growth factor 1 receptor and focal adhesion kinase in VSMCs remain to be studied.

Epidermal growth factor (EGF-receptor “trans”-activation by ROS

Recently it has become apparent that the EGF receptor is a significant contributor in a number of signaling networks activated by stimuli that do not directly interact with the receptor (“transactivation”) (10). These stimuli include various GPCR

TABLE 1. ROS-SENSITIVE TYROSINE KINASES IN VSMCS

| Tyrosine kinase | Agonists | References |
|-----------------|--|------------------------|
| Receptor | | |
| EGF receptor | H ₂ O ₂ , AngII, PDGF, oxidized LDL | 25, 28, 70, 76, 84, 93 |
| PDGF receptor | H ₂ O ₂ , AngII, PDGF, oxidized LDL | 21, 33, 74, 86 |
| FGF receptor | H ₂ O ₂ | 71 |
| Non-receptor | | |
| JAK2 | H ₂ O ₂ , O ₂ ^{•-} , AngII, thrombin | 28, 55, 56, 78 |
| PYK2 | H ₂ O ₂ , AngII | 24 |
| Src | H ₂ O ₂ , AngII | 74, 93 |

FGF, fibroblast growth factor.

ligands, membrane depolarizing agents, and environmental stressors that rapidly induce EGF receptor tyrosine phosphorylation (some act within 1 min). In particular, studies have shown that not only AngII (16) but also H₂O₂ (70), endothelin (40), thrombin (44), oxidized LDL (84), β -migrating very-LDL (99), and mechanical stretch (41) transactivate the EGF receptor in cultured VSMCs (Fig. 1). Moreover, a rapid transactivation of aortic EGF receptor by acute AngII infusion *in vivo* was reported (46). Since the EGF receptor transactivation mediates several critical mitogenic signal transductions by these stimuli, the EGF receptor could be a point of signal convergence by which several cardiovascular risk factors mediate pathophysiological responses associated with cardiovascular diseases. How is the EGF receptor transactivated by these risk factors in VSMCs?

H₂O₂ has been shown to induce tyrosine phosphorylation of the EGF receptor and to cause the association of Shc–Grb2–Sos complex with the EGF receptor in VSMCs (70). In VSMCs, the EGF receptor is a redox-sensitive tyrosine kinase that is activated by a GPCR agonist, AngII (25, 93). It has been reported that *N*-acetylcysteine, a potent ROS scavenger, is capable of inhibiting AngII-induced EGF receptor transactivation, and that H₂O₂ induced EGF receptor autophosphorylation at Tyr1,068, a Grb2 binding site, strongly suggesting ROS are required for EGF receptor transactivation induced by AngII (25). Furthermore, lysophosphatidic acid-induced transactivation of the EGF receptor requires ROS in HeLa cells (12). Taken together, these data indicate that the EGF receptor utilizes ROS for transactivation by GPCRs, however, the precise mechanisms by which ROS transactivate the EGF receptor are still unknown.

A mechanism by which ROS transactivate the EGF receptor may involve inhibition of tyrosine phosphatases, which in turn results in enhanced phosphorylation of tyrosine kinases (22). In support of this notion, H₂O₂ and various other thiol-oxidizing agents could inhibit dephosphorylation of the EGF receptor in rat-1 cells (47). However, in VSMCs there exists another mechanism by which ROS can transactivate the EGF receptor. Metalloprotease-dependent heparin-binding EGF-like growth factor (HB-EGF) generation has been implicated in EGF receptor transactivation initiated through several GPCRs

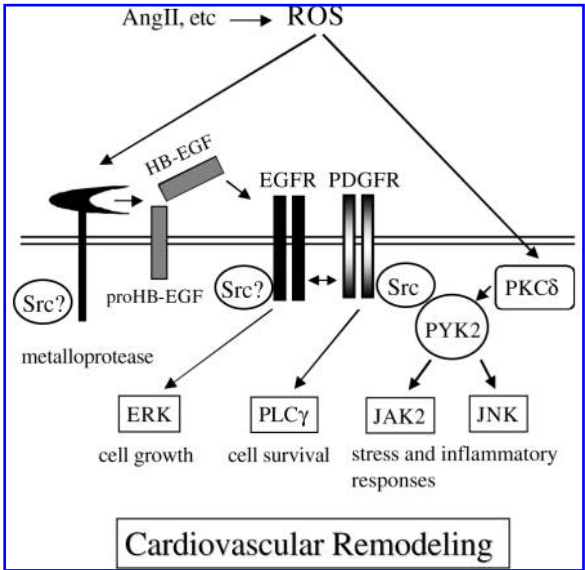


FIG. 2. Hypothetical signaling mechanisms of ROS lead to transactivation of the EGF receptor (EGFR) and PDGF receptor (PDGFR) and activation of non-receptor tyrosine kinases (PYK2, JAK2, and Src) promoting functional responses critical for vascular pathogenesis.

(19, 67). HB-EGF belongs to the EGF family of growth factors and is a strong stimulant of mitogenesis and migration of VSMCs, two critical events involved in the pathogenesis of atherogenic vascular diseases (68). In fact, HB-EGF is present in atherosclerotic plaques, and VSMCs produce HB-EGF in response to various pathogens (68), suggesting a critical role for HB-EGF in mediating vascular remodeling.

In VSMCs, H₂O₂ stimulates EGF receptor transactivation via metalloprotease-dependent HB-EGF generation (Fig. 2) (28). Although the metalloprotease responsible for the HB-EGF generation induced by ROS has not been identified, both matrix metalloproteases (87, 97) and a disintegrin and metalloprotease (ADAM) family of metalloproteases (5, 42, 51) have been implicated in ectodomain shedding of HB-EGF stimulated by various agonists. Interestingly, H₂O₂ was recently shown to enhance ADAM17 activity directly and ADAM17-mediated ectodomain shedding (98). Activation of metalloproteases is thought to occur via a thiol group from a cysteine residue within the inhibitory prodomain of these metalloproteases that interacts with zinc in their catalytic domain. Since ROS are known to interact with thiol groups, they may oxidize these electrophilic thiol groups and disrupt the cysteine–zinc bond, leading to activation of the metalloprotease.

In addition, the mechanism of ROS-induced EGF receptor transactivation may further involve intracellular Ca²⁺ elevation, upstream tyrosine kinases, and/or heterodimer formation with the PDGF receptor. Intracellular Ca²⁺ elevation is required for the EGF receptor transactivation by AngII, endothelin, and mechanical stretch in VSMCs (16, 40, 41). It has been reported that Ca²⁺ exists upstream of HB-EGF generation (19). Since H₂O₂ has no major effect on intracellular Ca²⁺ in VSMCs (24), Ca²⁺ may mediate ROS production, which leads to EGF

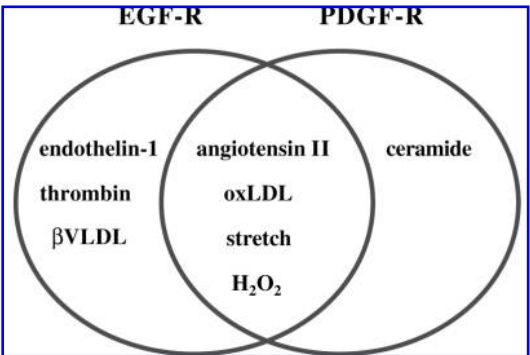


FIG. 1. Multiple risk factors lead to phosphorylation and transactivation of both the EGF receptor (EGF-R) and the PDGF receptor (PDGF-R) in VSMCs. βVLDL, β -migrating very-LDL; oxLDL, oxidized LDL.

receptor transactivation. c-Src has been implicated in EGF receptor transactivation by several GPCRs (4, 54, 93), and activation of Src-mediated cascades could further amplify ROS production in VSMCs (81). Moreover, H_2O_2 -induced EGF receptor transactivation was inhibited by a selective Src inhibitor, PP2, in endothelial cells (11). ROS may modulate c-Src, which then activates the metalloprotease responsible for HB-EGF generation through protein–protein interaction. In this regard, c-Src appears to exist upstream of HB-EGF release in fibroblasts (66). Alternatively, c-Src phosphorylates the EGF receptor in response to ROS to induce EGF receptor transactivation. To support this idea, only certain tyrosine sites have been shown to be phosphorylated by AngII in an ROS-dependent manner in VSMCs (93). Also, it has been shown that PDGF stimulation of VSMCs results in the formation of a PDGF- β receptor–EGF receptor heterodimer leading to transactivation of EGF receptor, which involves ROS production and a Src kinase activation (76). Therefore, c-Src may have multiple roles in mediating EGF receptor transactivation by H_2O_2 . In contrast to these studies, a recent paper has demonstrated that the direct association of EGF receptor and AngII receptor after AngII stimulation induces EGF receptor transactivation in COS cells, suggesting that an ROS-independent mechanism of EGF receptor transactivation may also exist in certain conditions (82).

Transactivation of the EGF receptor has been implicated in several disease processes (5, 51, 63), making it a current and important topic of signal transduction research. AngII-induced transactivation of the EGF receptor is necessary for the activation of downstream signal transduction molecules, such as ERK, p38 MAPK, Akt, and p70 S6 kinase, that subsequently promote hypertrophy and hyperplasia of VSMCs (15, 18, 19). Among these kinases, p38 MAPK and Akt have been shown to be redox-sensitive in VSMCs, whereas redox sensitivity of ERK in VSMCs remains controversial (23, 25, 91, 92). Interestingly, EGF receptor transactivation through GPCRs is required for cardiac hypertrophy induced by AngII as well as by pressure overload (5, 43). Also, EGF receptor transactivation mediates VSMC migration in response to AngII (75). Therefore, it is now becoming clear that the transactivation of the EGF receptor plays a significant role in the development and progression of cardiovascular diseases and that this signaling cascade may provide for alternative therapeutic targets for prevention of such diseases.

PDGF receptor “trans”-activation by ROS

The PDGF receptor, which exists as an α - or β -isoform, is a transmembrane spanning receptor tyrosine kinase (35). Similar to the EGF receptor, the PDGF receptor can be activated not only by its cognate ligands but also by other stimuli in a ligand-independent manner in VSMCs (Fig. 1). Cyclic mechanical stretch rapidly (within 4 min) induces tyrosine phosphorylation of PDGF- α receptor and its association with Grb2, an adaptor molecule essential for ERK/MAPK activation in VSMCs (37). Oxidized LDL, ceramide, and AngII transactivate the PDGF- β receptor in cultured VSMCs (21, 53). The transactivation of the PDGF- β receptor appears to involve ROS in VSMCs (21, 33). Since PDGF is a potent mitogen and chemoattractant in VSMCs and has long been implicated in

atherosclerosis as well as other cardiovascular diseases (35), these findings indicate the unique activation mechanism of the PDGF receptor induced by ROS and its downstream pathophysiological significance.

In VSMCs, H_2O_2 stimulates phosphorylation of the PDGF- β receptor on tyrosine residues, one of which was identified as Tyr1,021, a phospholipase C (PLC)- γ binding site (74). Both the binding of PLC- γ to phosphorylated Tyr1,021 in the C-terminal tail of the PDGF- β receptor and the activation of PLC- γ are believed to be involved in cell growth and chemotaxis in certain circumstances (34). The fact that PLC- γ is recruited to the PDGF- β receptor after H_2O_2 stimulation is important supportive evidence regarding H_2O_2 -induced PDGF- β receptor transactivation (74). Interestingly, in the same study, H_2O_2 induced association of c-Src and protein kinase C (PKC)- δ with the PDGF- β receptor. These non-receptor kinases (c-Src and PKC- δ) are required for H_2O_2 -induced PDGF- β receptor transactivation but not for PDGF-BB (ligand)-induced receptor activation. Thus the PDGF receptor transactivation is ligand-independent, and therefore does not require PDGF receptor kinase activity or metalloprotease activation (74). In addition, it has been shown that a non-receptor tyrosine kinase, PYK2, is required for PDGF receptor but not EGF receptor transactivation by ROS in VSMCs by using dominant-negative PYK2 mutants (20) (Fig. 3). Other investigators have demonstrated an involvement of phosphotyrosine phosphatase in ROS-induced PDGF- β receptor transactivation in mesangial cells (9) and in VSMCs (21). Alternatively, ROS-dependent transactivation of PDGF receptor induced by AngII may require a tyrosine kinase distinct from Src, PYK2, or JAK2 in VSMCs (33). It should be noted that extracellular administration of H_2O_2 and receptor stimulation of H_2O_2 production may differentially activate signaling pathways, which may explain some of the disagreement in the literature. Taken together, these findings clearly highlight the unique mechanism of PDGF receptor transactivation that is distinct from a ligand-dependent autophosphorylation of the PDGF receptor as well as

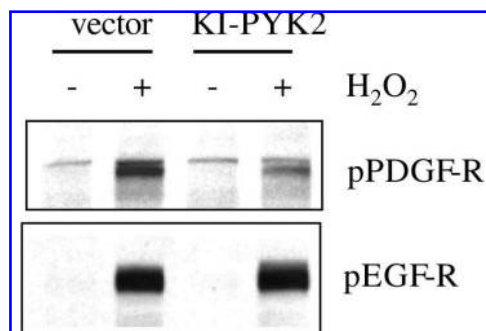


FIG. 3. ROS-dependent phosphorylation of the PDGF receptor requires tyrosine kinase activity of PYK2. VSMCs were transfected with either vector or kinase-inactive (KI)-PYK2 (10 multiplicity of infection) for 48 h and stimulated by H_2O_2 (20 μM) for 10 min. The cell lysates were immunoblotted with antibodies to Tyr1,021-phosphorylated PDGF receptor (pPDGF-R) and Tyr1,068-phosphorylated EGF receptor (pEGF-R) (25, 72). PDGF receptor but not EGF receptor phosphorylation was inhibited by KI-PYK2 transfection.

metalloprotease-dependent transactivation of the EGF receptor by ROS in VSMCs (Fig. 2).

In comparison with the EGF receptor, little is known about the downstream function of PDGF receptor transactivation. Since its transactivation mechanism is not via autophosphorylation, the downstream significance should be different from PDGF-mediated responses in VSMCs. Our finding that PLC- γ as well as c-Src and PKC- δ associates with the PDGF receptor suggests that the receptor acts as a scaffold in ROS signaling. Interestingly, PLC- γ activated by ROS was recently shown to be involved in cell survival against ROS-induced apoptosis (60). In addition, PDGF receptor phosphorylation was enhanced in balloon-injured carotid arteries, which was inhibited by AT₁ receptor antagonists (2). Taken together, these data suggest the possible involvement of PDGF receptor transactivation in the cardiovascular remodeling process.

JAK2 activation by ROS and its signaling

JAK2 is a member of the JAK family of tyrosine kinases, which are critical for signal transduction important for several biological functions. In particular, JAK activation is required for activation of the signal transducers and activators of transcription (STAT) pathway in response to activated cytokine receptors (69). An early study demonstrated that AngII could also stimulate tyrosine phosphorylation and activation of JAK2, which subsequently leads to STAT isoform tyrosine phosphorylation (57). Interestingly, AngII also stimulates the association of JAK2 with the AT₁ receptor in VSMCs (57), cardiac myocytes (48), and renal mesangial cells (58), which may partly explain the cytokine-like actions of AngII in mediating cardiovascular remodeling. In addition to AngII, a variety of other GPCR agonists have recently been shown to activate JAK/STAT pathways (94). Research has revealed that JAK2 activation by AngII requires ROS in VSMCs (28, 78) and that H₂O₂ rapidly and strongly induces JAK2 activation in VSMCs (28, 55). Similarly, H₂O₂ activate the JAK/STAT pathway in cultured fibroblasts (1, 83). These findings suggest that GPCR agonists activate JAK2 through ROS production.

In VSMCs, intracellular Ca²⁺ elevation and PKC- δ activation initiated by PLC-derived second messengers are involved in AngII-induced JAK2 activation. Furthermore, PYK2 is required for Ca²⁺- and PKC- δ -mediated JAK2 activation (27). These findings provide new information whereby GPCR-dependent JAK2 activation may be mediated by ROS. Recently, it has been reported that the PKC- δ isoform is required for JAK2 activation by H₂O₂ in VSMCs (28), which is in line with the above-mentioned finding. Previously, PKC- δ has been implicated in ROS-dependent activation of other tyrosine kinases such as c-Abl and c-Src (85). In this regard, several reports indicate that H₂O₂ stimulates PKC- δ activity in various cell types (49, 62). In fact, our own group has shown that H₂O₂ stimulates PKC- δ activity in VSMCs (28). Interestingly, H₂O₂-induced activation of PKC- δ is reported to be independent from tyrosine phosphatase inhibition (95). Utilizing dominant-negative PYK2 mutants, it has been shown that PYK2, which is also downstream of PKC- δ in VSMCs, is required for JAK2 activation but not for EGF receptor activation by H₂O₂ (26). In fibroblasts, a Src kinase was shown to exist upstream of JAK2 activation by ROS (1). As illustrated

in Fig. 2, these findings indicate that there are at least two major tyrosine kinase activation mechanisms utilized by ROS in VSMCs. One mechanism involves ROS-activated PKC- δ that leads to the activation of the PYK2/JAK2 pathway or the PDGF receptor transactivation. The other mechanism involves activation of a ROS-dependent metalloprotease cleavage of proHB-EGF to generate active HB-EGF that leads to EGF receptor transactivation.

Although the exact cellular function of JAK2 activation via ROS in VSMCs is unknown, JAK2 activation by AngII has been proposed to mediate VSMC proliferation (59). Also, the AT₁ receptor-mediated JAK/STAT pathway was shown to be involved in cardiac hypertrophy (64) and neointima formation after balloon injury (80). Along with the findings that ROS-dependent JAK2 activation is required for AngII-induced cytokine induction (78) and thrombin-induced heat shock protein induction in VSMCs (56), ROS-induced JAK2 activation could mediate cellular remodeling through a cytokine like inflammatory response in cardiovascular diseases.

PYK2 as unique ROS target

PYK2 is a non-receptor tyrosine kinase, also identified as cell adhesion kinase β , related adhesion focal tyrosine kinase, or calcium-dependent tyrosine kinase (6, 52, 77). PYK2 requires Ca²⁺ and/or PKC for its activation, which results from a wide variety of extracellular stimuli such as GPCR agonists, growth factors, cytokines, and environmental stresses (6, 52). In cultured VSMCs, it has been demonstrated that AngII rapidly stimulates PYK2 kinase activity (26, 73) and phosphorylation at Tyr402, a putative autophosphorylation site of PYK2 (24, 26).

In VSMCs, PYK2 is activated by extracellular administration of H₂O₂ (24). Depending on the cell type, PYK2 activates a downstream MAPK family that include ERK, c-Jun N-terminal kinase (JNK), and p38 MAPK (52, 65, 89). In VSMCs and cardiac fibroblasts, PYK2 is involved in AngII-induced JNK activation (26, 61). In addition, AngII stimulates association of PYK2 with c-Src in VSMCs (17), and c-Src was shown to mediate ROS-dependent JNK activation but not p38 MAPK or ERK activation in VSMCs (96). Some researchers have suggested a partial involvement of PYK2 in EGF receptor transactivation in fibroblasts (4), and that it plays a significant role in mediating AngII-induced growth-promoting signals (72). However, based on the above-mentioned findings, it is more likely that a ROS-sensitive kinase, PYK2, plays a major role in mediating JNK and JAK/STAT pathway activation leading to stress and inflammatory responses in VSMCs.

Src family kinase activation by ROS

Src family kinases now include nine members, of which Src, Fyn, and Yes are expressed in most tissues. These kinases can be activated by a variety of receptors, channels, and extracellular stresses, including ROS (88). As mentioned above, Src kinases seem to be critically involved in ROS-mediated activation of other tyrosine kinases such as activation of the EGF receptor, the PDGF receptor, and JAK2 in VSMCs. Importantly, c-Src was previously shown to mediate the phosphorylation of paxillin that is responsible for focal adhesion formation in VSMCs (39). Src kinase activation by ROS likely requires an interaction with PKC- δ . H₂O₂ induces phosphory-

lation of c-Src at Tyr418, a critical site for activation, leading to association of c-Src with PKC- δ in VSMCs. Also, a Src inhibitor blocks tyrosine phosphorylation of PKC- δ in response to H_2O_2 (74). These findings further suggest that interaction of PKC- δ with a non-receptor tyrosine kinase leads to their phosphorylation by each kinase toward the other, initiating ROS-dependent signal transduction (85).

It should be noted that many of the previous findings defined the role of Src kinases by using pharmacological inhibitors and dominant negative mutants, which could not establish the specific role of each isoform in mediating a specific cellular function. The recent development of isoform-specific knock-out cell lines may provide better information to elucidate the previous confusing role of ROS-dependent Src family kinase activation. In this regard, c-Src is proposed to exist upstream of JNK and Big MAP kinase (BMK1/ERK5) (3, 96), whereas Fyn mediates the JAK2 and Ras/MAPK/ERK cascade in mouse fibroblast cell lines (1). However, whether these cascades can be applied in VSMCs need further study.

FUTURE DIRECTION AND PERSPECTIVE

In summary, the findings discussed here clearly support the original hypothesis, which has been expanded to a new theory that multiple risk factors activate tyrosine kinases by ROS through distinct mechanisms in VSMCs and that each ROS-sensitive kinase has a unique role in mediating cardiovascular disease. Even though tremendous progress has been made in defining the role of ROS in tyrosine kinase-dependent signal transduction in cardiovascular systems, there is still a considerable void in our knowledge regarding the detailed mechanism of ROS-initiated signaling. Here, several mechanisms utilized by ROS to activate receptor and non-receptor type tyrosine kinases have been discussed, of which there are two significant contributors, a metalloprotease and PKC- δ . However, further extensive research is required to determine: (1) the identity of the metalloprotease that could be a critical ROS-sensing molecule in the EGF receptor transactivation pathway, (2) its activation mechanism, and (3) what's the downstream consequence *in vivo*. Further study is also needed to determine whether PKC- δ is directly activated by ROS, or whether there another molecule that senses ROS directly.

The long-term goal ideally is to be able to target these ROS-sensitive mechanisms with selective drugs to alleviate the pathophysiological conditions they promote. Based on this idea, you can imagine that we could target, for example, the stress and inflammatory or cell growth pathway independently according to the type of disease. Therefore, further characterization and understanding of the cellular mechanisms involved in ROS signal transduction via tyrosine kinase activation will provide new targets for effective therapies that will better enable us to control cardiovascular diseases.

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ABBREVIATIONS

ADAM, a disintegrin and metalloprotease; AngII, angiotensin II; AT₁, AngII type 1; EGF, epidermal growth factor; ERK, extracellular signal-regulated kinase; GPCR, G protein-coupled receptor; H_2O_2 , hydrogen peroxide; HB-EGF, heparin binding EGF-like growth factor; JNK, c-Jun N-terminal kinase; LDL, low density lipoprotein; MAPK, mitogen-activated protein kinase; PDGF, platelet-derived growth factor; PKC, protein kinase C; PLC, phospholipase C; ROS, reactive oxygen species; STAT, signal transducers and activators of transcription; VSMC, vascular smooth muscle cell.

REFERENCES

1. Abe J and Berk BC. Fyn and JAK2 mediate Ras activation by reactive oxygen species. *J Biol Chem* 274: 21003–21010, 1999.
2. Abe J, Deguchi J, Matsumoto T, Takuwa N, Noda M, Ohno M, Makuuchi M, Kurokawa K, and Takuwa Y. Stimulated activation of platelet-derived growth factor receptor *in vivo* in balloon-injured arteries: a link between angiotensin II and intimal thickening. *Circulation* 96: 1906–1913, 1997.
3. Abe J, Takahashi M, Ishida M, Lee JD, and Berk BC. c-Src is required for oxidative stress-mediated activation of big mitogen-activated protein kinase 1. *J Biol Chem* 272: 20389–20394, 1997.
4. Andreev J, Galisteo ML, Kranenburg O, Logan SK, Chiu ES, Okigaki M, Cary LA, Moolenaar WH, and Schlessinger J. Src and Pyk2 mediate G-protein-coupled receptor activation of epidermal growth factor receptor (EGFR) but are not required for coupling to the mitogen-activated protein (MAP) kinase signaling cascade. *J Biol Chem* 276: 20130–20135, 2001.
5. Asakura M, Kitakaze M, Takashima S, Liao Y, Ishikura F, Yoshinaka T, Ohmoto H, Node K, Yoshino K, Ishiguro H, Asanuma H, Sanada S, Matsumura Y, Takeda H, Beppu S, Tada M, Hori M, and Higashiyama S. Cardiac hypertrophy is inhibited by antagonism of ADAM12 processing of HB-EGF: metalloproteinase inhibitors as a new therapy. *Nat Med* 8: 35–40, 2002.
6. Avraham H, Park SY, Schinkmann K, and Avraham S. RAFTK/Pyk2-mediated cellular signalling. *Cell Signal* 12: 123–133, 2000.
7. Berk BC. Angiotensin II signal transduction in vascular smooth muscle: pathways activated by specific tyrosine kinases. *J Am Soc Nephrol* 10(Suppl): S62–S68, 1999.
8. Berk BC. Redox signals that regulate the vascular response to injury. *Thromb Haemost* 82: 810–817, 1999.
9. Callsen D, Sandau KB, and Brune B. Nitric oxide and superoxide inhibit platelet-derived growth factor receptor

- phosphotyrosine phosphatases. *Free Radic Biol Med* 26: 1544–1553, 1999.
10. Carpenter G. Employment of the epidermal growth factor receptor in growth factor-independent signaling pathways. *J Cell Biol* 146: 697–702, 1999.
11. Chen K, Vita JA, Berk BC, and Keaney JF, Jr. c-Jun N-terminal kinase activation by hydrogen peroxide in endothelial cells involves SRC-dependent epidermal growth factor receptor transactivation. *J Biol Chem* 276: 16045–16050, 2001.
12. Cunnick JM, Dorsey JF, Standley T, Turkson J, Kraker AJ, Fry DW, Jove R, and Wu J. Role of tyrosine kinase activity of epidermal growth factor receptor in the lysophosphatidic acid-stimulated mitogen-activated protein kinase pathway. *J Biol Chem* 273: 14468–14475, 1998.
13. Dostal DE and Baker KM. The cardiac renin-angiotensin system: conceptual, or a regulator of cardiac function? *Circ Res* 85: 643–650, 1999.
14. Dzau VJ. Mechanism of protective effects of ACE inhibition on coronary artery disease. *Eur Heart J* 19: J2–J6, 1998.
15. Eguchi S and Inagami T. Signal transduction of angiotensin II type 1 receptor through receptor tyrosine kinase. *Regul Peptides* 91: 13–20, 2000.
16. Eguchi S, Numaguchi K, Iwasaki H, Matsumoto T, Yamakawa T, Utsunomiya H, Motley ED, Kawakatsu H, Owada KM, Hirata Y, Marumo F, and Inagami T. Calcium-dependent epidermal growth factor receptor transactivation mediates the angiotensin II-induced mitogen-activated protein kinase activation in vascular smooth muscle cells. *J Biol Chem* 273: 8890–8896, 1998.
17. Eguchi S, Iwasaki H, Inagami T, Numaguchi K, Yamakawa T, Motley ED, Owada KM, Marumo F, and Hirata Y. Involvement of PYK2 in angiotensin II signaling of vascular smooth muscle cells. *Hypertension* 33: 201–206, 1999.
18. Eguchi S, Iwasaki H, Motley ED, Frank GD, Ueno H, Eguchi K, Marumo F, Hirata Y, and Inagami T. Intracellular signaling of angiotensin II-induced p70 S6 kinase phosphorylation at Ser411 in vascular smooth muscle cells: possible requirement of EGF receptor, RAS, ERK, and AKT. *J Biol Chem* 274: 36843–36852, 1999.
19. Eguchi S, Dempsey PJ, Frank GD, Motley ED, and Inagami T. Activation of MAP kinases by angiotensin II in vascular smooth muscle cells: metalloprotease-dependent EGF receptor activation is required for ERK and p38 MAP kinase, but not for JNK. *J Biol Chem* 276: 7957–7962, 2001.
20. Eguchi S, Frank GD, Saito S, and Inagami T. Distinct signal transduction mechanisms of EGF receptor and PDGF receptor transactivation by reactive oxygen species in vascular smooth muscle cells. *Circulation* 106: II-284, 2002.
21. Escargueil-Blanc I, Salvayre R, Vacaressse N, Jurgens G, Darblade B, Arnal JF, Parthasarathy S, and Negre-Salvayre A. Mildly oxidized LDL induces activation of platelet-derived growth factor beta-receptor pathway. *Circulation* 104: 1814–1821, 2001.
22. Finkel T. Redox-dependent signal transduction. *FEBS Lett* 476: 52–54, 2000.
23. Frank GD, Eguchi S, Yamakawa T, Tanaka S-i, Inagami T, and Motley ED. Involvement of reactive oxygen species in the activation of tyrosine kinase and extracellular signal-regulated kinase by angiotensin II. *Endocrinology* 141: 3120–3126, 2000.
24. Frank GD, Motley ED, Inagami T, and Eguchi S. PYK2/CAKbeta represents a redox-sensitive tyrosine kinase in vascular smooth muscle cells. *Biochem Biophys Res Commun* 270: 761–765, 2000.
25. Frank GD, Eguchi S, Inagami T, and Motley ED. N-Acetylcysteine inhibits angiotensin II-mediated activation of extracellular signal-regulated kinase and epidermal growth factor receptor. *Biochem Biophys Res Commun* 280: 1116–1119, 2001.
26. Frank GD, Eguchi S, Motley ED, Sasaki T, and Inagami T. Unique regulation of c-Jun N-terminal kinase by PYK2/CAK-beta in angiotensin II-stimulated vascular smooth muscle cells. *Biochem Biophys Res Commun* 286: 692–696, 2001.
27. Frank GD, Saito S, Motley ED, Sasaki T, Ohba M, and Inagami T. Requirement of Ca²⁺ and PKCdelta for Janus kinase 2 activation by angiotensin II: involvement of PYK2. *Mol Endocrinol* 16: 367–377, 2002.
28. Frank GD, Mifune M, Inagami T, Ohba M, Sasaki T, Higashiyama S, Dempsey PJ, and Eguchi S. Distinct mechanisms of receptor and non-receptor tyrosine kinase activation by reactive oxygen species in vascular smooth muscle cells: role of metalloprotease and protein kinase C-delta. *Mol Cell Biol* 23: 1581–1589, 2003.
29. Goodfriend TL, Elliott ME, and Catt KJ. Angiotensin receptors and their antagonists. *N Engl J Med* 334: 1649–1654, 1996.
30. Griendling KK and Ushio-Fukai M. Reactive oxygen species as mediators of angiotensin II signaling. *Regul Peptides* 91: 21–27, 2000.
31. Griendling KK, Ushio-Fukai M, Lassegue B, and Alexander RW. Angiotensin II signaling in vascular smooth muscle. New concepts. *Hypertension* 29: 366–373, 1997.
32. Griendling KK, Sorescu D, Lassegue B, and Ushio-Fukai M. Modulation of protein kinase activity and gene expression by reactive oxygen species and their role in vascular physiology and pathophysiology. *Arterioscler Thromb Vasc Biol* 20: 2175–2183, 2000.
33. Heeneman S, Haendeler J, Saito Y, Ishida M, and Berk BC. Angiotensin II induces transactivation of the platelet-derived growth factor beta receptor. *J Biol Chem* 275: 15926–15932, 2000.
34. Heldin CH. Simultaneous induction of stimulatory and inhibitory signals by PDGF. *FEBS Lett* 410: 17–21, 1997.
35. Heldin CH and Westermark B. Mechanism of action and in vivo role of platelet-derived growth factor. *Physiol Rev* 79: 1283–1316, 1999.
36. Heldin CH, Eriksson U, and Ostman A. New members of the platelet-derived growth factor family of mitogens. *Arch Biochem Biophys* 398: 284–290, 2002.
37. Hu Y, Bock G, Wick G, and Xu Q. Activation of PDGF receptor alpha in vascular smooth muscle cells by mechanical stress. *FASEB J* 12: 1135–1142, 1998.
38. Irani K. Oxidant signaling in vascular cell growth, death, and survival: a review of the roles of reactive oxygen species in smooth muscle and endothelial cell mitogenic and apoptotic signaling. *Circ Res* 87: 179–183, 2000.
39. Ishida T, Ishida M, Suero J, Takahashi M, and Berk BC. Agonist-stimulated cytoskeletal reorganization and signal

- transduction at focal adhesions in vascular smooth muscle cells require c-Src. *J Clin Invest* 103: 789–797, 1999.
40. Iwasaki H, Eguchi S, Ueno H, Marumo F, and Hirata Y. Endothelin-mediated vascular growth requires p42/p44 mitogen-activated protein kinase and p70 S6 kinase cascades via transactivation of epidermal growth factor receptor. *Endocrinology* 140: 4659–4668, 1999.
 41. Iwasaki H, Eguchi S, Ueno H, Marumo F, and Hirata Y. Mechanical stretch stimulates growth of vascular smooth muscle cells via epidermal growth factor receptor. *Am J Physiol* 278: H521–H529, 2000.
 42. Izumi Y, Hirata M, Hasuwa H, Iwamoto R, Umata T, Miyado K, Tamai Y, Kurisaki T, Sehara-Fujisawa A, Ohno S, and Mekada E. A metalloprotease-disintegrin, MDC9/meltrin-g/ADAM9 and PKC δ are involved in TPA-induced ectodomain shedding of membrane-anchored heparin-binding EGF-like growth factor. *EMBO J* 17: 7260–7272, 1998.
 43. Kagiya S, Eguchi S, Frank GD, Inagami T, Zhang YC, and Phillips MI. Angiotensin II-induced cardiac hypertrophy and hypertension are attenuated by epidermal growth factor receptor antisense. *Circulation* 106: 909–912, 2002.
 44. Kalmes A, Vesti BR, Daum G, Abraham JA, and Clowes AW. Heparin blockade of thrombin-induced smooth muscle cell migration involves inhibition of epidermal growth factor (EGF) receptor transactivation by heparin-binding EGF-like growth factor. *Circ Res* 87: 92–98, 2000.
 45. Kamata H and Hirata H. Redox regulation of cellular signalling. *Cell Signal* 11: 1–14, 1999.
 46. Kim S, Zhan Y, Izumi Y, Yasumoto H, Yano M, and Iwao H. In vivo activation of rat aortic platelet-derived growth factor and epidermal growth factor receptors by angiotensin II and hypertension. *Arterioscler Thromb Vasc Biol* 20: 2539–2545, 2000.
 47. Knebel A, Rahmsdorf HJ, Ullrich A, and Herrlich P. Dephosphorylation of receptor tyrosine kinases as target of regulation by radiation, oxidants or alkylating agents. *EMBO J* 15: 5314–5325, 1996.
 48. Kodama H, Fukuda K, Pan J, Makino S, Sano M, Takahashi T, Hori S, and Ogawa S. Biphasic activation of the JAK/STAT pathway by angiotensin II in rat cardiomyocytes. *Circ Res* 82: 244–250, 1998.
 49. Konishi H, Tanaka M, Takemura Y, Matsuzaki H, Ono Y, Kikkawa U, and Nishizuka Y. Activation of protein kinase C by tyrosine phosphorylation in response to H₂O₂. *Proc Natl Acad Sci U S A* 94: 11233–11237, 1997.
 50. Kunsch C and Medford RM. Oxidative stress as a regulator of gene expression in the vasculature. *Circ Res* 85: 753–766, 1999.
 51. Lemjabbar H and Basbaum C. Platelet-activating factor receptor and ADAM10 mediate responses to *Staphylococcus aureus* in epithelial cells. *Nat Med* 8: 41–46, 2002.
 52. Lev S, Moreno H, Martinez R, Canoll P, Peles E, Musacchio JM, Plowman GD, Rudy B, and Schlessinger J. Protein tyrosine kinase PYK2 involved in Ca(2+)-induced regulation of ion channel and MAP kinase functions. *Nature* 376: 737–745, 1995.
 53. Linseman DA, Benjamin CW, and Jones DA. Convergence of angiotensin II and platelet-derived growth factor receptor signaling cascades in vascular smooth muscle cells. *J Biol Chem* 270: 12563–12568, 1995.
 54. Luttrell LM, Della RG, van BT, Luttrell DK, and Lefkowitz RJ. Gbetagamma subunits mediate Src-dependent phosphorylation of the epidermal growth factor receptor. A scaffold for G protein-coupled receptor-mediated Ras activation. *J Biol Chem* 272: 4637–4644, 1997.
 55. Madamanchi NR, Li S, Patterson C, and Runge MS. Reactive oxygen species regulate heat-shock protein 70 via the JAK/STAT pathway. *Arterioscler Thromb Vasc Biol* 21: 321–326, 2001.
 56. Madamanchi NR, Li S, Patterson C, and Runge MS. Thrombin regulates vascular smooth muscle cell growth and heat shock proteins via the JAK-STAT pathway. *J Biol Chem* 276: 18915–18924, 2001.
 57. Marrero MB, Schieffer B, Paxton WG, Heerdt L, Berk BC, Delafontaine P, and Bernstein KE. Direct stimulation of Jak/STAT pathway by the angiotensin II AT1 receptor. *Nature* 375: 247–250, 1995.
 58. Marrero MB, Paxton WG, Schieffer B, Ling BN, and Bernstein KE. Angiotensin II signalling events mediated by tyrosine phosphorylation. *Cell Signal* 8: 21–26, 1996.
 59. Marrero MB, Schieffer B, Li B, Sun J, Harp JB, and Ling BN. Role of Janus kinase/signal transducer and activator of transcription and mitogen-activated protein kinase cascades in angiotensin II- and platelet-derived growth factor-induced vascular smooth muscle cell proliferation. *J Biol Chem* 272: 24684–24690, 1997.
 60. Martindale JL and Holbrook NJ. Cellular response to oxidative stress: signaling for suicide and survival. *J Cell Physiol* 192: 1–15, 2002.
 61. Murasawa S, Matsubara H, Mori Y, Masaki H, Tsutsumi Y, Shibasaki Y, Kitabayashi I, Tanaka Y, Fujiyama S, Koyama Y, Fujiyama A, Iba S, and Iwasaka T. Angiotensin II initiates tyrosine kinase Pyk2-dependent signalings leading to activation of Rac1-mediated c-Jun NH₂-terminal kinase. *J Biol Chem* 275: 26856–26863, 2000.
 62. Ohmori S, Shirai Y, Sakai N, Fujii M, Konishi H, Kikkawa U, and Saito N. Three distinct mechanisms for translocation and activation of the delta subspecies of protein kinase C. *Mol Cell Biol* 18: 5263–5271, 1998.
 63. Pai R, Soreghan B, Szabo IL, Pavelka M, Baatar D, and Tarnawski AS. Prostaglandin E₂ transactivates EGF receptor: a novel mechanism for promoting colon cancer growth and gastrointestinal hypertrophy. *Nat Med* 8: 289–293, 2002.
 64. Pan J, Fukuda K, Kodama H, Makino S, Takahashi T, Sano M, Hori S, and Ogawa S. Role of angiotensin II in activation of the JAK/STAT pathway induced by acute pressure overload in the rat heart. *Circ Res* 81: 611–617, 1997.
 65. Pandey P, Avraham S, Kumar S, Nakazawa A, Place A, Ghanem L, Rana A, Kumar V, Majumder PK, Avraham H, Davis RJ, and Kharbada S. Activation of p38 mitogen-activated protein kinase by PYK2/related adhesion focal tyrosine kinase-dependent mechanism. *J Biol Chem* 274: 10140–10144, 1999.
 66. Pierce KL, Tohgo A, Ahn S, Field ME, Luttrell LM, and Lefkowitz RJ. Epidermal growth factor (EGF) receptor-dependent ERK activation by G protein-coupled receptors: a co-culture system for identifying intermediates upstream and downstream of heparin-binding EGF shedding. *J Biol Chem* 276: 23155–23160, 2001.

67. Prenzel N, Zwick E, Daub H, Leserer M, Abraham R, Wallasch C, and Ullrich A. EGF receptor transactivation by G-protein-coupled receptors requires metalloproteinase cleavage of proHB-EGF. *Nature* 402: 884–888, 1999.
68. Raab G and Klagsbrun M. Heparin-binding EGF-like growth factor. *Biochim Biophys Acta* 1333: F179–F199, 1997.
69. Rane SG and Reddy EP. Janus kinases: components of multiple signaling pathways. *Oncogene* 19: 5662–5679, 2000.
70. Rao GN. Hydrogen peroxide induces complex formation of SHC-Grb2-SOS with receptor tyrosine kinase and activates Ras and extracellular signal-regulated protein kinases group of mitogen-activated protein kinases. *Oncogene* 13: 713–719, 1996.
71. Rao GN. Protein tyrosine kinase is required for oxidant-induced extracellular signal-regulated protein kinase activation and c-fos and c-jun expression. *Cell Signal* 9: 181–187, 1997.
72. Rocic P and Lucchesi PA. Down-regulation by antisense oligonucleotides establishes a role for the proline-rich tyrosine kinase PYK2 in angiotensin II-induced signaling in vascular smooth muscle. *J Biol Chem* 276: 21902–21906, 2001.
73. Sabri A, Govindarajan G, Griffin TM, Byron KL, Samarel AM, and Lucchesi PA. Calcium- and protein kinase C-dependent activation of the tyrosine kinase PYK2 by angiotensin II in vascular smooth muscle. *Circ Res* 83: 841–851, 1998.
74. Saito S, Frank GD, Mifune M, Ohba M, Utsunomiya H, Motley ED, Inagami T, and Eguchi S. Ligand-independent trans-activation of the platelet-derived growth factor receptor by reactive oxygen species requires protein kinase C-delta and c-Src. *J Biol Chem* 277: 44695–44700, 2002.
75. Saito S, Frank GD, Motley ED, Dempsey PJ, Utsunomiya H, Inagami T, and Eguchi S. Metalloprotease inhibitor blocks angiotensin II-induced migration through inhibition of epidermal growth factor receptor transactivation. *Biochem Biophys Res Commun* 294: 1023–1029, 2002.
76. Saito Y, Haendeler J, Hojo Y, Yamamoto K, and Berk BC. Receptor heterodimerization: essential mechanism for platelet-derived growth factor-induced epidermal growth factor receptor transactivation. *Mol Cell Biol* 21: 6387–6394, 2001.
77. Sasaki H, Nagura K, Ishino M, Tobioka H, Kotani K, and Sasaki T. Cloning and characterization of cell adhesion kinase beta, a novel protein-tyrosine kinase of the focal adhesion kinase subfamily. *J Biol Chem* 270: 21206–21219, 1995.
78. Schieffer B, Luchtefeld M, Braun S, Hilfiker A, Hilfiker-Kleiner D, and Drexler H. Role of NAD(P)H oxidase in angiotensin II-induced JAK/STAT signaling and cytokine induction. *Circ Res* 87: 1195–1201, 2000.
79. Schmidt-Ott KM, Kagiya S, and Phillips MI. The multiple actions of angiotensin II in atherosclerosis. *Regul Peptides* 93: 65–67, 2000.
80. Seki Y, Kai H, Shibata R, Nagata T, Yasukawa H, Yoshimura A, and Imaizumi T. Role of the JAK/STAT pathway in rat carotid artery remodeling after vascular injury. *Circ Res* 87: 12–18, 2000.
81. Seshiah PN, Weber DS, Rocic P, Valppu L, Taniyama Y, and Griendling KK. Angiotensin II stimulation of NAD(P)H oxidase activity: upstream mediators. *Circ Res* 91: 406–413, 2002.
82. Seta K and Sadoshima J. Phosphorylation of tyrosine 319 of the angiotensin II type 1 receptor mediates angiotensin II-induced trans-activation of the epidermal growth factor receptor. *J Biol Chem* 278: 9019–9026, 2003.
83. Simon AR, Rai U, Fanburg BL, and Cochran BH. Activation of the JAK-STAT pathway by reactive oxygen species. *Am J Physiol* 275: C1640–C1652, 1998.
84. Suc I, Meilhac O, Lajoie-Mazenc I, Vandaele J, Jurgens G, Salvayre R, and Negre-Salvayre A. Activation of EGF receptor by oxidized LDL. *FASEB J* 12: 665–671, 1998.
85. Sun X, Frank W, Datta R, Kharbanda S, and Kufe D. Interaction between protein kinase C delta and the c-Abl tyrosine kinase in the cellular response to oxidative stress. *J Biol Chem* 275: 7470–7473, 2000.
86. Sundareshan M, Yu ZX, Ferrans VJ, Irani K, and Finkel T. Requirement for generation of H₂O₂ for platelet-derived growth factor signal transduction. *Science* 270: 296–299, 1995.
87. Suzuki M, Raab G, Moses MA, Fernandez CA, and Klagsbrun M. Matrix metalloproteinase-3 releases active heparin-binding EGF-like growth factor by cleavage at a specific juxtamembrane site. *J Biol Chem* 272: 31730–31737, 1997.
88. Thomas SM and Brugge JS. Cellular functions regulated by Src family kinases. *Annu Rev Cell Dev Biol* 13: 513–609, 1997.
89. Tokiwa G, Dikic I, Lev S, and Schlessinger J. Activation of Pyk2 by stress signals and coupling with JNK signaling pathway. *Science* 273: 792–794, 1996.
90. Touyz RM and Schiffrin EL. Signal transduction mechanisms mediating the physiological and pathophysiological actions of angiotensin II in vascular smooth muscle cells. *Pharmacol Rev* 52: 639–672, 2000.
91. Ushio FM, Alexander RW, Akers M, and Griendling KK. p38 mitogen-activated protein kinase is a critical component of the redox-sensitive signaling pathways activated by angiotensin II. Role in vascular smooth muscle cell hypertrophy. *J Biol Chem* 273: 15022–15029, 1998.
92. Ushio-Fukai M, Alexander RW, Akers M, Yin Q, Fujio Y, Walsh K, and Griendling KK. Reactive oxygen species mediate the activation of Akt/protein kinase B by angiotensin II in vascular smooth muscle cells. *J Biol Chem* 274: 22699–22704, 1999.
93. Ushio-Fukai M, Griendling KK, Becker PL, Hilenski L, Halleran S, and Alexander RW. Epidermal growth factor receptor transactivation by angiotensin II requires reactive oxygen species in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 21: 489–495, 2001.
94. Williams JG. Serpentine receptors and STAT activation: more than one way to twin a STAT. *Trends Biochem Sci* 24: 333–334, 1999.
95. Yamamoto T, Matsuzaki H, Konishi H, Ono Y, and Kikkawa U. H₂O₂-induced tyrosine phosphorylation of protein kinase cdelta by a mechanism independent of inhibition of protein-tyrosine phosphatase in CHO and COS-7 cells. *Biochem Biophys Res Commun* 273: 960–966, 2000.
96. Yoshizumi M, Abe J, Haendeler J, Huang Q, and Berk BC. Src and Cas mediate JNK activation but not ERK1/2 and

- p38 kinases by reactive oxygen species. *J Biol Chem* 275: 11706–11712, 2000.
97. Yu WH, Woessner JF Jr, McNeish JD, and Stamenkovic I. CD44 anchors the assembly of matrilysin/MMP-7 with heparin-binding epidermal growth factor precursor and ErbB4 and regulates female reproductive organ remodeling. *Gene Dev* 16: 307–323, 2002.
98. Zhang Z, Oliver P, Lancaster JJ, Schwarzenberger PO, Joshi MS, Cork J, and Kolls JK. Reactive oxygen species mediate tumor necrosis factor alpha-converting, enzyme-dependent ectodomain shedding induced by phorbol myristate acetate. *FASEB J* 15: 303–305, 2001.
99. Zhao D, Letterman J, and Schreiber BM. β -Migrating very low density lipoprotein (β VLDL) activates smooth muscle mitogen-activated protein (MAP) kinase via G protein-coupled receptor-mediated transactivation of the epidermal growth factor (EGF) receptor. *J Biol Chem* 276: 30579–30588, 2001.

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2. Maria E. Lönn, Joanne M. Dennis, Roland Stocker. 2012. Actions of “antioxidants” in the protection against atherosclerosis. *Free Radical Biology and Medicine* **53**:4, 863-884. [[CrossRef](#)]
3. James E. Klaunig, Zemin Wang, Xinzhu Pu, Shaoyu Zhou. 2011. Oxidative stress and oxidative damage in chemical carcinogenesis. *Toxicology and Applied Pharmacology* . [[CrossRef](#)]
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5. P. R. Nagareddy, F. L. Chow, L. Hao, X. Wang, T. Nishimura, K. M. MacLeod, J. H. McNeill, C. Fernandez-Patron. 2009. Maintenance of adrenergic vascular tone by MMP transactivation of the EGFR requires PI3K and mitochondrial ATP synthesis. *Cardiovascular Research* **84**:3, 368-377. [[CrossRef](#)]
6. Jacqueline M. Lafky, Jason A. Wilken, Andre T. Baron, Nita J. Maihle. 2008. Clinical implications of the ErbB/epidermal growth factor (EGF) receptor family and its ligands in ovarian cancer. *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer* **1785**:2, 232-265. [[CrossRef](#)]
7. Julie K. Spix, Edward Y. Chay, Ethan R. Block, Jes K. Klarlund. 2007. Hepatocyte growth factor induces epithelial cell motility through transactivation of the epidermal growth factor receptor. *Experimental Cell Research* **313**:15, 3319-3325. [[CrossRef](#)]
8. Mazen Kurdi, George W Booz. 2007. Can the Protective Actions of JAK-STAT in the Heart be Exploited Therapeutically? Parsing the Regulation of Interleukin-6-Type Cytokine Signaling. *Journal of Cardiovascular Pharmacology* **50**:2, 126-141. [[CrossRef](#)]
9. Timothy M. Millar, Van Phan, Lee Anne Tibbles. 2007. ROS generation in endothelial hypoxia and reoxygenation stimulates MAP kinase signaling and kinase-dependent neutrophil recruitment. *Free Radical Biology and Medicine* **42**:8, 1165-1177. [[CrossRef](#)]
10. K COOPER, K LIU, L HUDSON. 2007. Contributions of reactive oxygen species and mitogen-activated protein kinase signaling in arsenite-stimulated hemeoxygenase-1 production. *Toxicology and Applied Pharmacology* **218**:2, 119-127. [[CrossRef](#)]
11. C ZHOU, Z QIAN, S ZHENG, M XIANG. 2006. ERK1/2 pathway is involved in the inhibitory effect of crocetin on angiotensin II-induced vascular smooth muscle cell proliferation. *European Journal of Pharmacology* **535**:1-3, 61-68. [[CrossRef](#)]
12. Haruhiko Ohtsu , Gerald D. Frank , Hirotohi Utsunomiya , Satoru Eguchi . 2005. Redox-Dependent Protein Kinase Regulation by Angiotensin II: Mechanistic Insights and Its Pathophysiology. *Antioxidants & Redox Signaling* **7**:9-10, 1315-1326. [[Abstract](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
13. Rhian M. Touyz . 2005. Reactive Oxygen Species as Mediators of Calcium Signaling by Angiotensin II: Implications in Vascular Physiology and Pathophysiology. *Antioxidants & Redox Signaling* **7**:9-10, 1302-1314. [[Abstract](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
14. Gerald D. Frank , Satoru Eguchi , Evangeline D. Motley . 2005. The Role of Reactive Oxygen Species in Insulin Signaling in the Vasculature. *Antioxidants & Redox Signaling* **7**:7-8, 1053-1061. [[Abstract](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
15. Yuichiro J. Suzuki , Hiroko Nagase , Kai Nie , Ah-Mee Park . 2005. Redox Control of Growth Factor Signaling: Recent Advances in Cardiovascular Medicine. *Antioxidants & Redox Signaling* **7**:5-6, 829-834. [[Abstract](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]

16. Rhian M Touyz, Guoying Yao, Ernesto L Schiffrin. 2005. Role of the actin cytoskeleton in angiotensin II signaling in human vascular smooth muscle cells. *Canadian Journal of Physiology and Pharmacology* **83**:1, 91-97. [[CrossRef](#)]
17. Yuichiro J. Suzuki , Kathy K. Griendling . 2003. Redox Control of Growth Factor Signaling in Heart, Lung, and Circulation. *Antioxidants & Redox Signaling* **5**:6, 689-690. [[Citation](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]